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# Metabolizable protein requirements for maintenance and gain of growing goats

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#### Abstract

A database of 349 treatment mean observations, representing 3404 goats from 73 publications between 1973 and 2003, was used to determine metabolizable protein (MP) requirements for maintenance (MP<sub>m</sub>) and growth (MP<sub>g</sub>) of goats. Published CP degradation properties of feedstuffs and proportions of dietary ingredients were used to estimate MP intake (MPI, g/day), which was regressed against ADG, with both variables scaled by  $BW^{0.75}$ . Goats were classified as meat (>50% Boer; 60 observations), dairy (selected for milk production; 129 observations) and indigenous (160 observations) biotypes. Because of differences (P < 0.01) among biotypes in slopes, separate regressions were initially performed—meat: MPI = 2.55(S.E. =  $(0.360) + (0.441(S.E. = 0.0276) \times ADG)$  (n = 58;  $R^2 = 0.82$ ); dairy: MPI = 2.83(S.E. = 0.344) + (0.299(S.E. = 0.364)) 0.0238 × ADG) (n = 123;  $R^2 = 0.57$ ); and indigenous: MPI = 3.23 (S.E. = 0.212) + (0.281 (S.E. = 0.0304) × ADG) (n = 152;  $R^2 = 0.36$ ). Intercepts did not differ among biotypes (P = 0.37), but the slope for meat goats differed (P < 0.01) from those for dairy and indigenous goats; therefore, data sets for dairy and indigenous goats were pooled and split into development (n = 150) and evaluation (n = 125) subsets. Using the equation derived from the development data subset for dairy and indigenous goats (i.e., MPI =  $3.14(S.E. = 0.189) + (0.285(S.E. = 0.0168) \times ADG)$  (n = 144;  $R^2 = 0.67$ ), MPI for the evaluation subset was predicted; regressing observed against predicted MPI of the evaluation data subset resulted in an intercept and slope not different from 0 and 1, respectively (P > 0.05). The equation from the development subset for dairy and indigenous goats was compared with the equation from the meat goat data set; there was a difference (P < 0.01) in slopes but not in intercepts (P = 0.25). Therefore, a dummy variable (D = 1 for meat goats and 0 otherwise) was used to develop a common intercept equation: MPI =  $3.07(S.E. = 0.165) + (0.290(S.E. = 0.0150) \times ADG) + (0.114(S.E. = 0.0162) \times D \times ADG) (n = 0.0162) \times D \times ADG$ 202;  $R^2 = 0.75$ ). In conclusion, based on regression of MPI against ADG, MP<sub>m</sub> was 3.07 g/kg BW<sup>0.75</sup> for all biotypes of growing goats, and MPg was 0.404 and 0.290 g/g ADG for meat and other (dairy and indigenous) goats, respectively. © 2004 Elsevier B.V. All rights reserved.

Keywords: Goat; Metabolizable protein; Maintenance; Growth

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### 1. Introduction

Goats are important livestock for food and economic securities, particularly in developing countries of the world. However, there has been relatively less research on requirements of goats for nutrients such as protein than for other livestock species. It is now generally accepted that to best address protein needs of ruminants, feed protein reaching the small intestine intact and microbial protein synthesized in the rumen both should be considered, along with adjustment for the extent degradation in the small intestine. In accordance, metabolizable protein (MP) systems are now in common use for various classes of livestock (INRA, 1989; Wilkerson et al., 1993; AFRC, 1998; NRC, 2001), but such systems have not yet been extensively studied with goats. AFRC (1998) proposed an MP requirement for maintenance (MP<sub>m</sub>) of 2.19 g/kg BW<sup>0.75</sup> and an efficiency of MP conversion to net tissue protein gain of 59% for goats, but these estimates were based on findings with cattle and sheep. Therefore, objectives of this study were to determine  $MP_m$  and the MP requirement for gain  $(MP_g)$ for growing goats based on a database of treatment mean observations from publications of goat feeding and nutrition experiments in the literature.

#### 2. Materials and methods

#### 2.1. Database construction and data derivation

The database for growing goats (postweaning to 18 months of age) consisted of 349 treatment mean observations from 73 publications (Appendix), representing 3404 goats. Observations were categorized into three biotypes, meat ( $\geq$ 50% Boer), dairy (e.g., Saanen, Alpine, Damascus, Norwegian, Swedish Landrace and dairy crossbred) and indigenous (neither dairy nor meat, without Angora goats). The length of experiments ranged from 21 to 256 days, with an average of 87. Reports included information necessary to determine mean BW, BW gain or ADG, DM intake, ME intake, CP intake, ME and CP concentrations in the diet and dietary ingredient proportions.

A parallel database of CP degradation properties for ingredients in diets of the reports of this study was constructed (Table 1) to estimate MP intake (MPI), as

described by Nsahlai et al. (2004). Briefly, CP degradation properties included soluble CP (SolP), soluble non-protein N or CP as a percentage of SolP (SolNP), insoluble protein that can be potentially degraded (slowly, relative to soluble true protein) in the rumen and is available for digestion in the small intestine (SDP), rate of degradation of SDP (Rate<sub>SDP</sub>) and acid detergent insoluble protein (ADIP: indigestible in the rumen and intestines). SolP is described by NRC (2001) as non-protein N assumed to be instantly degraded in the rumen and true protein that rapidly escapes from in situ bags because of high solubility or very small particle size, comparable to the quickly degraded CP fraction of AFRC (1993). Soluble true protein was estimated as the difference between SolP and SolNP. The SDP fraction is comparable to the B fraction listed by AFRC (1993) and NRC (2001; presented in tabular form). A fraction of insoluble protein not subject to ruminal degradation but potentially degraded in the small intestine (rumen undegraded but intestinally digestible dietary protein; RUDDP) was calculated as the difference between total CP and the sum of SolP, SDP and ADIP. Feedstuff CP degradation properties were primarily derived from NRC (2000) for SolNP; NRC (2001) and AFRC (1993) for SolP, SDP and Ratespp: and NRC (2001), AFRC (1993) and NRC (2000) for ADIP, with an additional small number of listings derived from other sources. To calculate ADIP, it was assumed that all CP from urea in urea-treated wheat straw was soluble in acid detergent solution. Dietary levels of the different CP fractions and Rate<sub>SDP</sub> were based on CP degradation properties of feedstuffs and their dietary proportions.

Level of feeding (*L*) was ME intake divided by the ME requirement for maintenance (ME<sub>m</sub>). An ME<sub>m</sub> (kJ/kg BW<sup>0.75</sup>) was assumed based on AFRC (1998) recommendations for the net energy for maintenance requirement of 315 kJ/kg BW<sup>0.75</sup> and efficiency of ME utilization for maintenance ( $k_m =$ 0.503 + 0.019 × ME, MJ/kg DM). Energy costs for activity were not considered. SolNP was assumed completely degraded in the rumen (AFRC, 1993); thus, the extent of ruminal degradation of SolNP (ExSolNP) was equal to SolNP. Because it has not been clearly established how rates of digesta passage from the rumen of goats compare with other ruminant species, the extent of ruminal protein digestion was based in part on estimates of fluid and

Table 1							
Crude prote	in degradability	properties	for feedstuff	s used to	calculate	metabolizable	protein intake

Feedstuff	CP (g/g DM)	SolP (g/g CP) <sup>a</sup>	SDP (g/g CP) <sup>b</sup>	$\frac{k_{\rm d}}{({\rm h}^{-1})^{\rm c}}$	ADIP (g/g CP) <sup>d</sup>	RUDDP (g/g CP) <sup>e</sup>	NPCP (g/g SolP) <sup>f</sup>	Source <sup>g</sup>
Acacia macracantha	0.224	0.30	0.62	0.030	0.021	0.07	0.90	Kaitho et al. (1998)
leaves	0.221	0.50	0.02	0.050	0.021	0.07	0.90	Runno et ul. (1990)
Alfalfa hav	0.183	0.25	0.65	0.290	0.180	0	0.92	AFRC (1993)
Alfalfa meal	0.192	0.28	0.66	0.067	0.125	0	1.00	NRC (2001)
Ammonium chloride	1.642	1.00	0.00		0.000	0	1.00	NRC (2000)
Apple pomace	0.063	0.25	0.61	0.035	0.315	0	1.00	Ahn et al. (2002)
Barley grain	0.138	0.25	0.70	0.350	0.050	0	0.29	AFRC (1993)
Barley hay	0.125	0.22	0.60	0.080	0.093	0.09	0.93	AFRC (1993)
Bermudagrass hav	0.100	0.37	0.52	0.080	0.088	0.03	0.25	NRC (2001)
Blood meal	0.955	0.10	0.61	0.019	0.010	0.28	0.00	NRC (2001)
Bone meal	0.132	0.18	0.48	0.072	0.000	0.34	0.00	NRC (2000)
Brewers spent grain	0.292	0.18	0.65	0.047	0.120	0.05	0.75	NRC (2001)
Casava chips (tapioca)	0.038	0.25	0.70	0.120	0.050	0	0.45	NRC (2000)
Casava peel silage	0.058	0.14	0.39	0.070	0.080	0.39	1.00	Baah et al. (1999)
Chickpea	0.266	0.12	0.82	0.071	0.010	0.05	0.23	Hadiipanaviotou (2002)
Chickpea straw	0.085	0.30	0.35	0.053	0.177	0.17	0.00	NRC (2001)
Coconut meal	0.213	0.28	0.65	0.087	0.030	0.04	0.75	NRC (2001)
Concentrate	0.137	0.58	0.30	0.063	0.022	0.09	0.45	Ahn et al. (2002)
Corn grain (ground)	0.094	0.24	0.73	0.049	0.032	0	0.73	NRC (2001)
Corn bran	0.119	0.45	0.49	0.070	0.005	0.06	0.80	AFRC (1993)
Corn gluten meal	0.650	0.04	0.91	0.023	0.046	0.01	0.75	NRC (2001)
Corn stalks	0.041	0.20	0.66	0.040	0.136	0	0.95	NRC (2000)
Cottonseed meal	0.449	0.26	0.56	0.068	0.040	0.15	0.40	NRC (2001)
Cottonseed hulls	0.041	0.30	0.35	0.053	0.177	0.17	0.00	NRC (2001)
Dried beet pulp	0.100	0.05	0.91	0.020	0.060	0	0.96	NRC (2001)
Faba bean seed	0.314	0.67	0.33	0.039	0.020	0	0.23	NRC (2001)
Flemingia macrophylla leaves	0.112	0.13	0.13	0.002	0.017	0.73	0.96	Kaitho et al. (1998)
Gliricidia leaves	0.183	0.29	0.45	0.074	0.250	0.01	1.00	Ash (1990)
Groundnut hulls	0.078	0.23	0.76	0.050	0.087	0	0.00	AFRC (1993)
Groundnut cake	0.518	0.62	0.70	0.161	0.021	0	0.23	NRC (2001)
Guatemala grass	0.087	0.23	0.68	0.020	0.089	0	0.25	Moheni et al. (1996)
Guinea grass	0.094	0.06	0.57	0.073	0.089	0.28	0.25	Sampath et al. (1989)
Krishnachura leaves	0.145	0.18	0.60	0.028	0.200	0.02	0.92	Fleischer et al. (1998)
Leucaena leaves	0.269	0.30	0.62	0.030	0.021	0.07	0.90	Kaitho et al. (1993)
Lupin seed	0.345	0.30	0.67	0.261	0.034	0	0.68	NRC (2001)
Meat meal	0.576	0.35	0.40	0.060	0.032	0.22	0.27	NRC (2001)
Molasses	0.085	0.74	0.26	0.032	0.000	0.22	1.00	NRC (2001)
Mulberry (Morus alba) leaves	0.208	0.20	0.50	0.026	0.200	0.1	0.92	Liu et al. (2000)
Mustard oil cake	0.385	0.23	0.70	0.104	0.063	0	0.65	NRC (2001)
Nanier grass	0.078	0.46	0.52	0.110	0.002	Ő	0.02	NRC (2000)
Oat hav	0.078	0.35	0.52	0.043	0.103	0.02	0.93	NRC (2001)
Orcharderass hav	0.130	0.25	0.69	0.110	0.061	0	0.96	NRC (2000)
Paragrass ( <i>Brachiaria</i> <i>mutica</i> ) hay	0.092	0.09	0.70	0.054	0.110	0.1	0.95	Kalbande and Thomas (1997)
Peanut hav	0.171	0.39	0.50	0.140	0.099	0.02	0.96	NRC (2001)
Rhodesgrass hav	0.066	0.28	0.53	0.050	0.167	0.02	0.96	NRC (2001)
Rice bran	0.155	0.33	0.49	0.050	0.026	0.16	0.80	NRC (2001)
Rice straw (NaOH treated)	0.035	0.36	0.37	0.067	0.292	0	0.95	Vadiveloo (2000)

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Feedstuff	CP (g/g DM)	SolP (g/g CP) <sup>a</sup>	SDP (g/g CP) <sup>b</sup>	$k_{\rm d}$ $({\rm h}^{-1})^{\rm c}$	ADIP (g/g CP) <sup>d</sup>	RUDDP (g/g CP) <sup>e</sup>	NPCP (g/g SolP) <sup>f</sup>	Source <sup>g</sup>
Rice straw (urea treated)	0.035	0.11	0.66	0.034	0.138	0.09	0.95	Ibrahim et al. (1989)
Rice straw	0.053	0.41	0.42	0.023	0.168	0	0.95	Negi et al. (1988)
Sesbania grandiflora leaves	0.208	0.11	0.79	0.183	0.100	0.01	0.96	Ash (1990)
Sorghum grain	0.116	0.19	0.79	0.055	0.086	0	0.33	NRC (2001)
Sorghum-sudan hay	0.094	0.28	0.53	0.050	0.128	0.06	0.95	NRC (2001)
Soybean meal	0.499	0.05	0.93	0.037	0.020	0	0.55	Hadjipanayiotou (2002)
Soybean meal, formaldehyde treated	0.471	0.07	0.93	0.028	0.000	0	0.55	Michalet-Doreau and Nozière (1998)
Sugar cane bagasse	0.030	0.03	0.49	0.049	0.315	0.17	0.96	Ørskov et al. (1980)
Urea	2.880	1.00	0.00		0.000	0	1.00	AFRC (1993)
Vetch bran	0.256	0.56	0.44	0.167	0.010	0	0.96	NRC (2001)
Wheat bran	0.173	0.34	0.63	0.200	0.081	0	0.75	NRC (2001)
Wheat straw	0.048	0.09	0.51	0.014	0.292	0.1	0.95	NRC (2001)

<sup>a</sup> Soluble CP (g/g total CP).

<sup>b</sup> Slowly degradable protein (g/g total CP).

<sup>c</sup> Rate of degradation of SDP ( $h^{-1}$ ).

<sup>d</sup> Acid detergent insoluble CP (g/g total CP).

<sup>e</sup> Ruminally undegraded but intexstinally digestible protein (g/g total CP). Calculated as the difference between total CP and the sum of SolP, SDP and ADIP.

<sup>f</sup> Non-protein CP (g/g SolP). Derived from NRC (2000).

<sup>g</sup> Source of degradability parameters except for NPCP.

particulate passage rates. Ruminal outflow rate of particulates  $(k_p)$  was estimated following the equation proposed by AFRC (1993):  $k_p = -0.024 + 0.179(1 - e^{(-0.278L)})$ . Based on data from Nsahlai et al. (1999), ruminal fluid dilution rate  $(k_1)$  was determined as a function of  $k_p$ :  $k_1 = (k_p - 0.0018)/0.360$ . With an approach similar to that of Ngwa et al. (2001), passage rates were used to determine the extent of ruminal degradation of SolTP (ExSolTP) and SDP (ExSDP):

$$ExSolTP = SolTP \times \frac{Rate_{SolTP}}{Rate_{SolTP} + k_{l}} \text{ and}$$
$$ExSDP = SDP \times \frac{Rate_{SDP}}{Rate_{SDP} + k_{p}}$$

where Rate<sub>SolTP</sub> is the rate of degradation of SolTP. In vitro ammonia accumulation (*y*) data of Brown et al. (1998) for casein were used to derive Rate<sub>SolTP</sub>:

$$y = 2.75(S.E. = 0.537) + (9.88(S.E. = 1.101))$$
  
  $\times (1 - e^{-0.084(S.E. = 0.0265)time})$   
 $(R^2 = 0.98, n = 7)$ 

Thus, Rate<sub>SolTP</sub> was 0.084. Undegraded SolTP and SDP were calculated by difference. Total undegraded protein in the rumen (RUDP) was obtained by summing undegraded SolTP, undegraded SDP and RUDDP, which was assumed to be 0.90 digestible postruminally (AFRC, 1993) to obtain digestible undegraded protein (DUDP).

AFRC (1993) assumed efficiencies of capture of N in ExSolNP and ExSolTP of 0.8 and in ExSDP of 1.0. Hence, effective ruminally degraded CP (ERDP) was the sum of  $0.8 \times \text{ExSolNP}$ ,  $0.8 \times \text{ExSolTP}$  and  $1.0 \times$ ExSDP. Furthermore, because utilization of ERDP in microbial CP synthesis depends on energy availability, energy from ruminal fermentation (RFE) was derived from listings in Appendix A of AFRC (1993) of ME and RFE concentrations in dietary concentrates and forages. Means of RFE were 92.6 (n = 11; S.D. = 4.35) and 82.0 (n = 18; S.E. = 2.75) of forage and concentrate ME (MJ/kg DM), respectively. These estimates were used along with ME intake and dietary concentrate and forage ME concentrations and proportions to estimate RFE (MJ/day). Using the equation proposed by AFRC (1993), microbial protein (MicP) was estimated for conditions with adequate ruminal availability of nitrogenous compounds as

MicP (g) = 
$$(7 + 6(1 - e^{(-0.35L)})) \times RFE$$

In accordance with ARC (1980), when the RFE-based estimate of MicP was greater than ERDP, ruminal availability of nitrogenous compounds was assumed limiting and, thus, MicP was set equal to ERDP. Assuming MicP N to be 0.25 nucleic acid N and that microbial true protein is 0.85 digestible, digestible microbial true protein (DMTP) was estimated as  $0.6375 \times \text{MicP}$  (AFRC, 1993). MPI was derived by adding DUDP and DMTP. A summary of important variables in the complete database for prediction of MP<sub>m</sub> and MP<sub>g</sub> is presented in Table 2.

#### 2.2. Regression analyses

MPI was regressed against ADG because objectives were to determine MP requirements for maintenance and growth rather than to determine efficiencies of MP use. In addition, it is desirable for the independent variable to be the one determined with greatest accuracy. MPI and ADG were scaled by kg BW<sup>0.75</sup>. Using PROC GLM of SAS (1990), differences among biotypes in intercepts and slopes of equations from regressions of MPI against biotype, ADG and their interaction were tested by analysis of covariance (Snedecor and Cochran, 1978). There was a difference among biotypes in slopes (P < 0.05), though not in intercepts (P = 0.86), suggesting the use of dummy variables to address the slope difference. However, because of the possibility that observations with relatively high residuals in data sets for each biotype might have contributed to the slope difference, separate regressions for each biotype were first performed.

Data for meat, dairy and indigenous goats consisted of 60, 129 and 160 treatment mean observations and represented 591, 1793 and 1019 goats, respectively. A summary of the data sets is presented in Table 3. Preceding simple linear regression, regressions of MPI against linear, quadratic and cubic effects of ADG were performed. Inclusion of quadratic and cubic ef-

Table 2

Summary of variables in the entire database (n = 349) for prediction of metabolizable protein requirements

Variable	Mean	S.D.	Min <sup>a</sup>	Max <sup>b</sup>
Soluble CP (g/g total CP)	0.267	0.1055	0.063	0.67
Soluble non-protein CP (g/g soluble CP)	0.63	0.2442	0.022	1
Slowly degradable protein (SDP; g/g total CP)	0.594	0.099	0.246	0.868
Rate of SDP degradation $(h^{-1})$	0.103	0.0651	0.024	0.35
Acid detergent insoluble CP (g/g total CP)	0.062	0.0349	0.01	0.227
DRUDP <sup>c</sup> (g/g total CP)	0.048	0.0568	0	0.411
Mean BW (kg)	21.8	9.34	5.5	52.2
Forage (% DM)	49	31.73	0	100
CP (% DM)	15.1	3.74	5.3	27.8
ME (MJ/kg DM)	10.2	1.75	3.6	14.4
DM intake (kg/day)	0.71	0.346	0.15	1.95
ME intake (MJ/day)	7.3	3.81	0.901	17.9
ADG (g/day)	101	77.8	-18	326
ADG $(g/(kg BW^{0.75} \times day))$	9.6	6.04	-2.4	28.1
CP intake (g/day)	110	63.4	13	312
RUDDP intake (g/day)	4.9	6.29	0	30.8
Total digestible ruminally undegraded protein intake <sup>d</sup> (g/day)	29	22.1	-2	102
Microbial CP (g/day)	57	33.9	6	157
Metabolizable protein intake (g/day)	66	41.8	2	203
Metabolizable protein intake (g/(kg BW <sup><math>0.75</math></sup> × day))	6.3	2.65	0.4	12.9

<sup>a</sup> Minimum.

<sup>b</sup> Maximum.

<sup>c</sup> Ruminally undegraded but intestinally digestible protein.

<sup>d</sup> Sum of RUDDP and ruminally undegraded soluble and insoluble true protein.

Summery of detabase subsets for prediction of metabolizable protein requirements for maintenance and gain of growing goats

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Variable	Meat goats <sup>a</sup> $(n = 60)$				Dairy go	$ats^b (n =$	129)		Indigenous goats <sup>c</sup> $(n = 160)$			
	Mean	S.D.	Min <sup>d</sup>	Max <sup>e</sup>	Mean	S.D.	Min	Max	Mean	S.D.	Min	Max
Mean BW (kg)	33.4	7.26	18.1	49	22.6	6.73	10.9	52.2	16.7	7.58	5.5	43
Forage (% DM)	50.3	17.97	12	74	40.6	35.67	0	100	55.2	31.03	0	100
CP (% DM)	16.3	2.04	10.2	23.6	15.3	3.47	8	20	14.4	4.28	5.3	27.8
ME (MJ/kg DM)	10.3	1.13	9.2	12.2	10.8	2	5.8	14.4	9.7	1.57	3.6	13.8
DM intake (kg/day)	1.1	0.39	0.47	1.95	0.77	0.265	0.34	1.41	0.52	0.23	0.15	1.45
MEI <sup>f</sup> (MJ/day)	10.9	3	5.7	17.9	8.5	3.83	3	17	5	2.3	0.9	14.4
CPI <sup>g</sup> (g/day)	178	63.4	71	312	120	55.2	34	258	76	43.4	13	255
ADG (g)	169	74.6	64	294	133	75.8	8	326	50	37	-18	222
ADG (g/kg BW <sup>0.75</sup> )	12.1	4.7	5.94	22.9	12.8	6.61	1.3	28.1	6.19	3.8	-2.4	18.7
MPI <sup>h</sup> (g/day)	112	41.2	47	202	73	38.4	22	173	42	24.8	2	164
MPI (g/kg BW <sup>0.75</sup> )	8	2.29	4.1	12.5	6.9	2.84	2.3	12.9	5.1	2.03	0.4	12.4

<sup>a</sup> Meat:  $\geq$ 50% Boer.

<sup>b</sup> Dairy: Saanen, Alpine, Damascus, Norwegian, Swedish Landrace and dairy crossbreed.

<sup>c</sup> Indigenous: neither meat nor dairy, without Angora goats.

<sup>d</sup> Minimum.

<sup>e</sup> Maximum.

f ME intake.

<sup>g</sup> Crude protein intake.

<sup>h</sup> Metabolizable protein intake.

fects of ADG resulted in small differences in  $R^2$  (i.e., 0.008, 0.018 and 0.038 for meat, dairy and indigenous goats, respectively) compared with simple linear regression. For meat and dairy goat data sets, linear, quadratic and cubic effects were not significant (P > 0.36 and 0.23, respectively), but the linear effect was significant (P < 0.01 and 0.05, respectively) when the cubic effect was removed from the model. For indigenous goats, quadratic and cubic effects were significant (P < 0.01), but the linear effect was not (P = 0.74); however, removal of the cubic effect resulted in a significant linear effect (P < 0.01). Therefore, simple linear regressions were performed.

For each regression analysis, the residual (difference between actual and predicted values) for each treatment mean observation was compared with various multiples of the residual S.D. (rS.D.). Observations with residual greater than selected rS.D. were removed, and changes in  $R^2$  and root mean square error (RMSE) were viewed. The rS.D. used to exclude observations was chosen on the basis of a moderate to appreciable increase in explained variability, with retention of the maximum number of observations. Observations removed were examined for possible reasons for high residuals (Chatterjee et al., 2000). Final equations were tested for differences among biotypes in intercepts and slopes. Intercepts did not differ (P = 0.37) among biotypes; the slope for dairy goats was similar (P = 0.64) to that for indigenous goats, but the slope for meat goats differed (P < 0.01) from those for dairy and indigenous goats. Hence, data sets for dairy and indigenous goats were pooled, and the data for meat goats were analyzed separately. The pooled data set for dairy and indigenous goats (i.e., non-meat goats), including 275 treatment means and representing 2673 goats, was split into development and evaluation subsets by report. Data in the two subsets were made as homogeneous as possible for most important variables (e.g., MPI, ADG and mean BW) by exchange of observations from a small number of reports. Mean, minimum and maximum values for important variables are summarized in Table 4. With the development subset, linear, quadratic and cubic effects of ADG on MPI were checked to justify the use of simple linear regression as described previously. The modified equation from the development subset was used to predict MPI in the evaluation subset. Observed MPI was regressed against predictions to determine whether the intercept and slope differed from 0 to 1, respectively.

A comparison of prediction equations for meat and non-meat goats revealed similar intercepts (P = 0.25)

Table 3

Table 4

Summary of development and evaluation subsets of pooled data set of dairy and indigenous goats for prediction of metabolizable protein requirements of growing goats

Variable	Development						Evaluation				
	n	Mean	S.D.	Min <sup>a</sup>	Max <sup>b</sup>	n	Mean	S.D.	Min	Max	
Mean BW (kg)	150	19.3	8.33	6.4	52.2	125	18.6	6.68	5.5	35.1	
DM intake (kg/day)	150	0.61	0.259	0.19	1.35	125	0.61	0.239	0.15	1.37	
CP (% DM)	150	14.6	3.82	5.3	23.7	125	14.7	4.02	6.9	23.6	
Forage (% DM)	150	46.8	35.84	0	100	125	52.6	31.9	0	100	
CP intake (g/day)	150	91	47.9	13	236	125	90	49.8	26	258	
ADG (g/day)	150	86	76.8	-18	326	125	83	62.8	0	294	
ADG $(g/(kg BW^{0.75} \times day))$	150	9	6.92	-2.4	28.1	125	9	5.4	0	25	
MP <sup>c</sup> intake (g/day)	150	54	31.7	2	147	125	52	29.3	13	173	
MP intake $(g/(kg BW^{0.75} \times day))$	150	5.7	2.48	0.4	12.1	125	5.7	2.21	2.3	12.2	
ME (MJ/kg DM)	150	10.2	1.95	3.6	14.4	125	10.1	1.7	5.8	12.7	
ME intake (MJ/day)	150	6.5	3.6	0.9	16.7	125	6.2	2.93	1.7	17	

<sup>a</sup> Minimum.

<sup>b</sup> Maximum.

<sup>c</sup> Metabolizable protein.

and a difference in slopes (P < 0.01); therefore, a dummy variable, D (D = 0 for non-meat goats, D = 1 for meat goats) was used to address the slope difference. The GLM model included ADG and the interaction of D and ADG. The final equation consisted of a common intercept for the three biotypes, a common slope for dairy and indigenous goats and a slope correction or adjustment term for meat goats. The intercept of the equation was considered the MP<sub>m</sub> and the slope MP<sub>g</sub>.

#### 3. Results

#### 3.1. Initial regressions

#### 3.1.1. Meat goats

The equation for the regression of MPI  $(g/kg^{0.75})$  against ADG  $(g/kg^{0.75})$  was

$$MPI = 2.82(S.E. = 0.401) + (0.428(S.E. = 0.0310) \times ADG)$$
  
(n = 60; R<sup>2</sup> = 0.77; RMSE = 1.118) (1)

Although the  $R^2$  of Eq. (1) was fairly high, removal of two observations with residuals greater than  $2.0 \times$ rS.D. yielded a slightly greater  $R^2$ :

MPI = 2.55(S.E. = 0.360)  
+ (0.441(S.E. = 0.0276) × ADG)  
(n = 58; 
$$R^2$$
 = 0.82; RMSE = 0.989) (2)

Regression lines for Eqs. (1) and (2) are presented in Fig. 1. The two excluded treatment mean observations were from the same study (Soto-Navarro et al., 2004) with Boer × Spanish wethers. Although reasons for high residuals are not apparent, these observations entailed use of diets containing corn gluten and fish meals that are relatively high in ruminally undegraded protein and ADG was low compared with others in the same study. Based on Eq. (2), preliminary estimates of MP<sub>m</sub> and MP<sub>g</sub> were 2.55 g/kg BW<sup>0.75</sup> and 0.441 g/g ADG, respectively.

# 3.1.2. Dairy goats

The equation for the regression of MPI  $(g/kg^{0.75})$  against ADG  $(g/kg^{0.75})$  was

MPI = 3.12(S.E. = 0.397)  
+ (0.295(S.E. = 0.0276) × ADG)  
(n = 129; 
$$R^2 = 0.47$$
; RMSE = 2.068) (3)

To improve fit of the model, six observations with residuals greater than 2.0 rS.D. were removed, and the resultant equation was



Fig. 1. Relationship between MP intake (MPI,  $g/kg^{0.75}$ ) and ADG ( $g/kg^{0.75}$ ) of growing meat goats. Points are observed values, the dashed line (---) represents the regression line for all observations in the data set and the solid line (---) is for the regression after removal of observations with high residuals (*x*: observations removed) and describes the equation: MPI = 2.55(S.E. = 0.360) + (0.441(S.E. = 0.0276) × ADG) (n = 58;  $R^2 = 0.82$ ). MBW = kg BW<sup>0.75</sup>.

$$MPI = 2.83(S.E. = 0.344)$$

+ (0.299(S.E. = 0.0238) × ADG)  
(
$$n = 123; R^2 = 0.57; RMSE = 1.778$$
) (4)

Regression lines for Eqs. (3) and (4) are presented in Fig. 2. The six excluded observations were above the regression line with relatively high MPI (approximately 154 g/day) and intermediate ADG (approximately 161 g/day). There were no apparent unique characteristics of these observations with respect to other variables, such as mean BW, dietary CP concentrations of CP or forage, ME intake, genotype, gender, etc. Preliminary estimates of MP<sub>m</sub> and MP<sub>g</sub> for growing dairy goats from Eq. (4) were 2.83 g/kg BW<sup>0.75</sup> and 0.299 g/g ADG, respectively.

#### 3.1.3. Indigenous goats

The equation for the regression of MPI  $(g/kg^{0.75})$  against ADG  $(g/kg^{0.75})$  was

MPI = 
$$3.19(S.E. = 0.253)$$
  
+ (0.306(S.E. = 0.0349) × ADG)  
(n = 160; R<sup>2</sup> = 0.33; RMSE = 1.672) (5)

To improve the fit of model, eight observations with residuals greater than 2.0 rS.D. were removed, yield-ing the following equation:

MPI = 3.23(S.E. = 0.212)  
+ (0.281(S.E. = 0.0304) × ADG)  
(n = 152; 
$$R^2 = 0.36$$
; RMSE = 1.356) (6)

Excluded observations did not seem to have commonalities in variables such as mean BW, breed, gender, dietary CP, ME or forage concentrations, ADG or MPI. Although Eq. (6) accounted for only 36% of variation, further deletion of observations with residuals greater than  $1.5 \times \text{rS.D.}$  resulted in a moderate increase in  $R^2$  (i.e., 0.44) but did not appreciably change the intercept (i.e., 3.22) or slope (i.e., 0.278) and markedly decreased the number of observations in the data set (i.e., 13% change). Therefore, Eq. (6) was considered most appropriate. Regression lines for Eqs. (5) and (6) are presented in Fig. 3. Based on Eq. (6), preliminary estimates of MP<sub>m</sub> and MP<sub>g</sub> for growing indigenous goats were  $3.23 \text{ g/kg BW}^{0.75}$  and 0.281 g/g ADG, respectively.



Fig. 2. Relationship between MP intake (MPI,  $g/kg^{0.75}$ ) and ADG ( $g/kg^{0.75}$ ) of growing dairy goats. Points are observed values, the dashed line (---) represents the regression line for all observations in the data set and the solid line (---) is for the regression after removal of observations with high residuals (*x*: observations removed) and describes the equation: MPI =  $2.83(S.E. = 0.344) + (0.299(S.E. = 0.0238) \times ADG)$  (n = 123;  $R^2 = 0.57$ ). MBW = kg BW<sup>0.75</sup>.



Fig. 3. Relationship between MP intake (MPI,  $g/kg^{0.75}$ ) and ADG ( $g/kg^{0.75}$ ) of growing indigenous goats. Points are observed values, the dashed line (---) represents the regression line for all observations in the data set and the solid line (---) is for the regression after removal of observations with high residuals (*x*: observations removed) and describes the equation: MPI =  $3.23(S.E. = 0.212) + (0.281(S.E. = 0.0304) \times ADG)$  (n = 152;  $R^2 = 0.36$ ). MBW = kg BW<sup>0.75</sup>.

#### 3.2. Comparison of equations

Because of the removal of a total of 16 observations, data sets resulting in Eqs. (2), (4) and (6) were combined and differences among biotypes in intercepts and slopes were tested. Slopes differed (P < 0.01) and intercepts were similar (P = 0.37). A paired comparison test indicated a similar (P = 0.64) slope between dairy and indigenous goats and a difference (P < 0.01) in slopes between meat and non-meat (dairy and indigenous) goats; therefore, meat goat data were analyzed separately and data for dairy and indigenous goats were pooled.

#### 3.3. $MP_m$ and $MP_g$ for non-meat goats

The pooled data set for non-meat goats (all observations) was split into development and evaluation subsets by report or publication. With the development subset, the linear effect of ADG on MPI was significant (P < 0.01), whereas quadratic (P = 0.39) and cubic (P = 0.31) effects were not; hence, a simple linear regression was conducted:

$$MPI = 3.12(S.E. = 0.206) + (0.282(S.E. = 0.0181) \times ADG) \\ (n = 150; R^2 = 0.62; RMSE = 1.529)$$
(7)

After removing six observations with residuals greater than 2.0 rS.D., the following modified equation was obtained:

MPI = 
$$3.14(S.E. = 0.189)$$
  
+ (0.285(S.E. = 0.0168) × ADG)  
(n = 144; R<sup>2</sup> = 0.67; RMSE = 1.396) (8)

Regression lines for Eqs. (7) and (8) are presented in Fig. 4. Five of the six observations removed had relatively low dietary CP and forage concentrations.

Using Eq. (8), the predicted MPI (MPI<sub>pred</sub>) was calculated for the evaluation subset. The regression of observed MPI against MPI<sub>pred</sub> resulted in this equa-



Fig. 4. Relationship between MP intake (MPI, g/kg<sup>0.75</sup>) and ADG (g/kg<sup>0.75</sup>) of development subset for growing dairy and indigenous goats. Points are observed values, the dashed line (---) represents the regression line for all observations in the data set and the solid line (—) is for the regression after removal of observations with high residuals (*x*: observations removed) and describes the equation: MPI =  $3.14(S.E. = 0.189) + (0.285(S.E. = 0.0168) \times ADG)$  (n = 144;  $R^2 = 0.67$ ). MBW = kg BW<sup>0.75</sup>.



Fig. 5. Relationship between MP intake (MPI,  $g/kg^{0.75}$ ) and ADG ( $g/kg^{0.75}$ ) of growing goats: ( $\bullet$ ) observations for growing meat goats; ( $\bigcirc$ ) observations for dairy and indigenous goats. The dashed line (---) describes the regression line for growing meat goats and the solid line (-) is for growing dairy and indigenous goats. The common intercept equation is: MPI =  $3.07(S.E. = 0.165) + (0.290(S.E. = 0.0150) \times ADG) + (0.114(S.E. = 0.0162) \times D \times ADG)$  (n = 202;  $R^2 = 0.75$ ). D = 1 for growing meat goats and 0 otherwise. MBW = kg BW<sup>0.75</sup>.

tion: MPI = 0.022(S.E. = 0.5509) + (0.994(S.E. = 0.0933) × MPI<sub>pred</sub>) (n = 125;  $R^2 = 0.48$ ). The intercept and slope of the equation were not different from 0 (P = 0.97) to 1 (P = 0.96), respectively. Thus, Eq. (8) provided unbiased estimates of MP<sub>m</sub> ( $3.14 \text{ g/kg BW}^{0.75}$ ) and MP<sub>g</sub> (0.285 g/g ADG) for non-meat goats.

# 3.4. Final equation for estimations of $MP_m$ and $MP_g$

Because of the removal of observations from the development data subset for non-meat goats, Eqs. (2) and (8) were tested for differences by analysis of covariance. There was a difference in slopes (P < 0.01) but not in intercepts (P = 0.25). Hence, a dummy variable D (D = 1 for meat goats and 0 otherwise) was used in the regression analysis. The common intercept equation from the regression of MPI against ADG was

$$MPI = 3.07(S.E. = 0.165)$$

+ (0.290(S.E. = 0.0150) × ADG)  
+ (0.114(S.E. = 0.0162) × 
$$D$$
 × ADG)  
( $n = 202; R^2 = 0.75; RMSE = 1.295$ ) (9)

Regression lines of Eq. (9) are presented in Fig. 5. Based on Eq. (9),  $MP_m$  for all growing goats was 3.07 g/kg BW<sup>0.75</sup> and MP<sub>g</sub> for meat goats and non-meat goats was 0.404 and 0.290 g/g ADG, respectively.

#### 4. Discussion

# 4.1. Derivation of CP degradation properties and MPI calculation

Because few CP degradation properties have been determined with goats, and it is likely that such characteristics do not differ appreciably among ruminant species, CP degradation properties for other ruminants were used to calculate MPI. The method of estimating MPI was quite similar to that of AFRC (1993), as well as the NRC (2000) Level 1 approach. CP degradation measures used were based on in situ ruminal N disappearance as discussed by Ørskov (1980), Ørskov and MacLeod (1982) and Ørskov and Shand (1997).

# 4.2. $MP_m$

In addition to determination of MP<sub>m</sub> by regression analysis, a factorial approach can also be used for comparison purposes. To do so, the net protein requirement for maintenance (NPm) was assumed to be the sum of endogenous urinary CP (EUCP), metabolic fecal CP (MFCP) and scurf CP (SCP) losses. The EUCP for goats from Luo et al. (2004) of  $1.031 \text{ g/kg BW}^{0.75}$ and the MFCP for goats from Moore et al. (2004) of 2.67% DM intake were used. In addition, the SCP estimate of  $0.2 \times BW^{0.60}$  from NRC (1984) was assumed. Using the mean BW (21.8 kg) and DM intake (0.711 kg/day) in the database of this study, NPm was 3.04 g/kg BW<sup>0.75</sup>. Assuming an efficiency of MP use for maintenance  $(k_{pm})$  of 1.00 (AFRC, 1993), the corresponding MP<sub>m</sub> was  $3.04 \text{ g/kg BW}^{0.75}$ , which is very close to the estimate from regression analysis in the present study  $(3.07 \text{ g/kg BW}^{0.75})$ . With a similar regression approach, Wilkerson et al. (1993) reported a slightly greater MPm of growing beef steers of  $3.8 \text{ g/kg BW}^{0.75}$ , which was adopted as the recommendation for beef cattle of NRC (2000). However, based on seven studies in the period of 1950-1980, NRC (1981) proposed a digestible CP requirement for maintenance of goats of 2.82 g/kg BW<sup>0.75</sup>, which based on common magnitudes of energy loss in urine equates to a slightly lower MP<sub>m</sub> than our estimate. The MP<sub>m</sub> requirement for goats recommended by AFRC (1998;  $2.19 \text{ g/kg BW}^{0.75}$ ), based on data with other species, was also lower. Likewise, based on N balance of male goats in one experiment, INRA (1989) suggested an  $MP_m$  of 2.30 g/kg BW<sup>0.75</sup>.

# 4.3. MPg

With the approach used to determine MP requirements, an inherent assumption is that MP intake limited growth. It is possible that for some observations ME intake was relatively more limiting than intake of MP, which would have contributed to variability in ADG not accounted for by MP intake. However, since ME intake is a primary determinant of MP intake through its influence on microbial protein synthesis, it is most likely that the degree to which ME intake might have been more limiting than MP intake, or vice versa, was small. Nonetheless, because the  $MP_g$  requirement was determined from change in MP intake per unit change in ADG, it seems appropriate to consider this  $MP_g$  requirement a maximum rather than average and, relatedly, that supplying additional MP as a safety factor to ensure desired levels of performance is unwarranted.

Factors responsible for the greater MPg estimate for meat goats than for dairy and indigenous goats are unclear; however, reports of a higher protein concentration in BW gain in meat goats could be involved (Mmbengwa et al., 2000). Many estimates of MPg have been derived by separate prediction of the protein concentration in empty or live BW gain (NRC, 1985, 2000, 2001; AFRC, 1993, 1998) and an assumed constant efficiency of MP use for growth  $(k_{tg})$ , resulting in a wide range of MPg. This method was not used in the present study because composition of gain was not reported in most publications and there is not currently available an accurate means of prediction for a wide array of goat genotypes and production systems. The MPg of 0.24 g/g ADG for all goats of AFRC (1998) based on this approach is somewhat lower than determined in the present study (i.e., 0.290 g/g ADG for non-meat goats and 0.404 g/g ADG for meat goats). This difference may be ascribed to factors such as method of determination, the assumption of AFRC (1998) for  $k_{tg}$ , experimental conditions, body composition, growth rate, etc. For example, protein concentration in tissue gain of beef cattle decreases, and that of fat increases, as growth rate and BW increase (Byers, 1982). However, our findings by regression analysis are in line with cattle studies in which MPg was assumed constant (INRA, 1989; Ainslie et al., 1993; Wilkerson et al., 1993). For example, Wilkerson et al. (1993), with a similar regression approach, noted an MPg for growing beef steers of 0.305 g/g ADG. Likewise, INRA (1989) reported an MP<sub>g</sub> value of 0.36 g/g ADG for all goats.

# 4.4. Efficiency of MP use for maintenance and growth of goats

Assumed efficiencies of MP use for maintenance  $(k_{pm})$  vary widely among protein systems (0.75 for ARC, 1980; 0.70 for SCARM, 1994; 1.00 for AFRC, 1998; 0.67 for NRC, 2001). To compare our estimate of MP<sub>m</sub> from regression analysis to that with a factorial approach, the  $k_{pm}$  assumed was 1.00, based on

the justification of AFRC (1992) and concepts outlined by Oldham (1987) regarding amino acid requirements of non-ruminants. NRC (2001) proposed a  $k_{pm}$ value of 0.67; however, the correction of MFCP by NRC (2001) for bacterial cell debris based on several assumptions would partially compensate for the lower  $k_{pm}$ .

Net protein gain is the multiple of ADG and composition of gain, with the latter influenced by growth rate, physiological maturity, previous nutrition, gender, etc. (NRC, 1985). Protein concentration in BW gain can be calculated assuming a concentration in empty BW gain such as 16% (AFRC, 1998) and conversion to a live BW basis by dividing by 1.09 (ARC, 1980), resulting in 147 g/kg. Also, with the equation of AFRC (1998) (protein concentration in BW gain, g/kg = $157.22 - [0.694 \times BW, kg]$ ) and mean BW (33.4 and 19.3 for meat and non-meat goats, respectively), protein concentration in BW gain was 134 and 144 g/kg for meat and non-meat goats, respectively. Efficiency of MP use for protein accretion  $(k_{tg})$  for growing meat and non-meat goats, respectively, was 0.36 and 0.51 from the first method and 0.33 and 0.50 from the second. The meat goat  $k_{tg}$  is lower than values of NRC (1985, 0.50), INRA (1989, 0.65), AFRC (1993, 1998, 0.59) and SCARM (1994, 0.70). Although it is possible that protein turnover is relatively high in meat goats, because MP<sub>m</sub> was similar among genotypes, this difference could involve a greater protein concentration in BW gain by meat goats than predicted by these two methods.

### 5. Summary

Using a database of treatment mean observations from reports with growing goats, along with feedstuff CP degradation properties, MP requirements were determined by regressing MPI against ADG. The MP requirement for maintenance was 3.07 g/kg BW<sup>0.75</sup>, and the MP requirement for BW gain of meat goats was 0.404 g/g ADG and that for dairy and indigenous goats was 0.290 g/g ADG. Because of the large number of observations on which these estimates are based, they seem useful in determining diet composition for growing goats as well as predicting performance. However, because MPI may not have in all instances been more limiting to growth than ME intake, it seems appropriate to consider the estimates as maximum requirements rather than averages and, relatedly, that supplying additional MPI as a safety factor is unwarranted.

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# Appendix

- Abate, A., Pfeffer, E., 1986. Changes in nutrient intake and performance by goats fed coffee pulp-based diets followed by a commercial concentrate. Anim. Feed Sci. Technol. 14, 1–10.
- Abdel-Rahman, K.M., El Kaschab, S., 1996. Nutritional performance of Damascus and Jamnapari crossbred goats. World Rev. Anim. Prod. 31, 55–58.
- Adejumo, J.O., 1995. Effect of legume supplements on cassava peel silage utilization by West African Dwarf goats. Trop. Agric. 72, 175–177.
- Adeloye, A.A., 1995. The value of cowpea husk to the goat. Biores. Technol. 52, 281–282.
- Adeloye, A.A., Yousouf, M.B., 2001. Influence of nickel supplementation from nickel sulphate hexahydrate and nickel–sodium monofluorophosphate on the performance of the West African dwarf kids. Small Rumin. Res. 39, 195–198.
- Ahmed Muna, M.M., Shafei Ammar, I.El., 2001. Effect of water and feed restriction on body weight change and nitrogen balance in desert goats fed high and low quality forages. Small Rumin. Res. 41, 19–27.
- Anandan, S., Sastry, V.R.B., Musalia, L.M., Agrawal, D.K., 1996. Growth rate and nutrient efficiency of growing goats fed urea ammoniated neem (*Azadirachta indica*) seed kernel meal as protein supplement. Small Rumin. Res. 22, 205–212.
- Animut, G., Merkel, R.C., Abebe, G., Sahlu, T., Goetsch, A.L., 2002. Effects of level of broiler litter in diets containing wheat straw on performance of Alpine doelings. Small Rumin. Res. 44, 125–133.
- Aregheore, E.M., 2002a. Chemical evaluation and digestibility of Cocoa (*Theobroma cacao*) byproducts fed to goats. Trop. Anim. Health Prod. 34, 339–348.

- Aregheore, E.M., 2002b. Voluntary intake and digestibility of fresh, wilted and dry Leucaena (*Leucaena leucocephala*) at four levels to a basal diet of guinea grass (*Panicum maximum*). Asian-Aust. J. Anim. Sci. 15, 1139–1146.
- Ash, A.J., Norton, B.W., 1987. Studies with the Australian cashmere goat. I. Growth and digestion in male and female goats given pelleted diets varying in protein content and energy level. Aust. J. Agric. Res. 38, 957–969.
- Ash, A.J., Petaia, L., Ako, H., 1992. Nutritional value of *Sesbania grandiflora* leaves for monogastrics and ruminants. Trop. Agric. 69, 223–228.
- Bamikole, M.A., Ezenwa, I., Akinsoyinu, A.O., Arigbede, M.O., Babayemi, O.J., 2001. Performance of West African dwarf goats fed Guinea grass-Verano stylo mixture, N-fertilized and unfertilized Guinea grass. Small Rumin. Res. 39, 145–152.
- Beede, D.K., Schelling, G.T., Mitchell Jr., G.E., Tucker, R.E., 1985. Utilization by growing goats of diets that contain monensin and low or excess crude protein: comparative slaughter experiment. J. Anim. Sci. 61, 1230–1242.
- Beede, D.K., Schelling, G.T., Mitchell Jr., G.E., Tucker, R.E., Gill, W.W., Koenig, S.E., Lindsey, T.O., 1986. Nitrogen utilization and digestibility by growing steers and goats of diets that contain monensin and low crude protein. J. Anim. Sci. 62, 857–863.
- Cameron, M.R., Hart, S.P., Sahlu, T., Gilchrist, C., Coleman, S.W., Goetsch, A.L., 2001. Effects of gender and age on performance and harvest traits of Boer × Spanish goats. J. Appl. Anim. Res. 20, 141–155.
- Cochran, R.C., Del Carpio, A., Parker, C.F., Hallford, D.M., Van Keuren, R.W., 1984. Growth response of Peruvian Criollo goats consuming varying levels of *Acacia macracantha*, *Leucaena leucocephala* and corn stalks. Nutr. Rep. Int. 29, 495– 503.
- Economides, S., Koumas, A., Georghiades, E., Hadjipanayiotou, M., 1990. The effect of barley–sorghum grain processing and form of concentrate mixture on the performance of lambs, kids and calves. Anim. Feed Sci. Technol. 31, 105–116.
- El-Hag, M.G., El Shargi, K.M., 1998. Chemical composition and feeding value of chopped date fronds

(CDF) as affected by urea and date syrup extract treatment. Asian-Aust. J. Anim. Sci. 11, 685–691.

- El Muola, I.H.A., Babiker, S.A., El Khidir, O.A., Ibrahim, S.E., 1999. Meat production from female goat kids compared with males. J. Agric. Sci. (Camb.) 133, 223–226.
- Galgal, K.K., Norton, B.W., 1991. The value of Copra meal expeller pellets as concentrate feed for weaner goats. In: Saithanoo, S., Norton, B.W. (Eds.), Goat Production in the Asian Humid Tropics. The Australian International Development Assistance Bureau, Hat Yai, Thailand, pp. 144–153.
- Gelaye, S., Amoah, E.A., 1991. Nutritive value of florigraze rhizoma peanut as an alternative leguminous forage for goats. Small Rumin. Res. 6, 131–139.
- Gelaye, S., Amoah, E.A., Guthrie, P., 1990. Performance of yearling goats fed alfalfa and *Florigraze rhizoma* peanut hay. Small Rumin. Res. 3, 353–361.
- Gipson, T., 2000. Meat buck performance test—1999. In: Proceedings of the 14th Annual Goat Field Day. Agricultural Research and Extension Program, Langston University, Langston, OK, pp. 123–124.
- Gipson, T., 2001. Meat buck performance test—2000. In: Proceedings of the 15th Annual Goat Field Day. Agricultural Research and Extension Program, Langston University, Langston, OK, pp. 41–44.
- Gipson, T., 2002. Meat buck performance test—2001. In: Proceedings of the 16th Annual Goat Field Day. Agricultural Research and Extension Program, Langston University, Langston, OK, pp. 102–106.
- Gipson, T., 2003. Meat buck performance test—2002. In: Proceedings of the 17th Annual Goat Field Day. Agricultural Research and Extension Program, Langston University, Langston, OK, pp. 97–105.
- Goetsch, A.L., Detweiler, G., Sahlu, T., Dawson, L.J., 2001. Effects of different management practices on preweaning and early postweaning growth of Alpine kids. Small Rumin. Res. 41, 109–116.
- Hadjipanayiotou, M., 2002. Replacement of soybean meal and barley grain by chickpeas in lamb and kid fattening diets. Anim. Feed Sci. Technol. 96, 103–109.
- Hadjipanayiotou, M., Koumas, A., Hadjigavriel, G., Antoniou, T., Photiou, A., Theodoridou, M., 1996. Feeding dairy ewes and goats and growing lambs and kids mixtures of protein supplements. Small Rumin. Res. 21, 203–211.

- Havrevoll, O., Rajbhandari, S.P., Eik, L.O., Nedkvitne, J.J., 1995. Effects of different energy levels during indoor rearing on performance of Norwegian dairy goats. Small Rumin. Res. 15, 231–237.
- Huston, J.E., Bales, K.W., Engdahl, B.S., 1998. Effects of breed, sex, and ration type on production of market kid goats. In: Sheep Goat Wool Mohair. Texas Agricultural Experimental Station, San Angelo, TX, pp. 16–17.
- Ivey, D.S., Owens, F.N., Sahlu, T., Teh, T.H., Claypool, P.L., Goetsch, A.L., 2000. Growth and cashmere production by Spanish goats consuming ad libitum diets differing in protein and energy levels. Small Rumin. Res. 35, 133–139.
- James, C.S., Chandran, K., 1975. Enquiry into the role of minerals in experimental urolithiasis in goats. Indian Vet. J. 52, 251–258.
- Jia, Z.H., Sahlu, T., Fernandez, J.M., Hart, S., Teh, T.H., 1995. Effects of dietary protein level on performance of Angora and cashmere-producing Spanish goats. Small Rumin. Res. 16, 113–119.
- Johnson, T.J., Gherardi, S.G., Dhaliwal, S., 1994. Diet quality effects the cashmere production and liveweight of Western Australian cashmere goats. Aust. J. Exp. Agric. 34, 1107–1112.
- Kibria, S.S., Nahar, T.N., Mia, M.M., 1994. Tree leaves as alternative feed resource for Black Bengal goats under stall-fed conditions. Small Rumin. Res. 13, 217–222.
- Louca, A., Economides, S., Hancock, J., 1977. Effects of castration on growth rate, feed conversion efficiency and carcass quality in Damascus goats. Anim. Prod. 24, 387–391.
- Louca, A., Hancock, J., 1977. Genotype by environment interactions for postweaning growth in the Damascus breed of goat. J. Anim. Sci. 44, 927–931.
- Louca, A., Papas, A., 1973. The effect of different proportions of carob pod meal in the diet on the performance of calves and goats. Anim. Prod. 17, 139–146.
- Lu, C.D., Potchoiba, M.J., 1990. Feed intake and weight gain of growing goats fed diets of various energy and protein levels. J. Anim. Sci. 68, 1751–1759.
- Luginbuhl, J.M., Poore, M.H., Conrad, A.P., 2000a. Effect of level of whole cottonseed on intake, digestibility, and performance of growing male goats fed hay-based diets. J. Anim. Sci. 78, 1677–1683.

- Luginbuhl, J.M., Poore, M.H., Spears, J.W., Brown, T.T., 2000b. Effect of dietary copper level on performance and copper status of growing meat goats. Sheep and Goat Res. J. 16, 65–71.
- Malamsha, P.C., Nuhikambele, V.R.M., Mtenga, L.A., 1999. White mulberry (*Morus alba*) as a potential feed supplement for stall-fed growing goats in highland areas of Tanzania. In: Mbaga, S.H., Ndemanisho, E.E., Kombe, R., Kakengi, A.M.V. (Eds.), Proceedings of the TSAP Scientific Conference Tanzania Society of Animal Production, Morogora, Tanzania, pp. 160–169.
- Mtenga, L.A., Kitaly, A.J., 1990. Growth performance and carcass characteristics of Tanzanian goats fed *Chloris gayana* hay with different levels of protein supplement. Small Rumin. Res. 3, 1–8.
- Ndemanisho, E.E., Mtenga, L.A., Kimbi, E.F.C., Kimambo, A.E., Mtengeti, E.J., 1998. Substitution of dry *Leucaena leucocephala* (DLL) leaves for cotton seed cake (CSC) as a protein supplement to urea-treated maize stover fed to dairy weaner goats. Anim. Feed Sci. Technol. 73, 365–374.
- Negesse, T., Rodehutscord, M., Pfeffer, E., 2001. The effect of dietary crude protein level on intake, growth, protein retention and utilization of growing male Saanen kids. Small Rumin. Res. 39, 243– 251.
- Njwe, R.M., 1990. Energy requirement of Cameroon dwarf goat. World Rev. Anim. Prod. 25, 61–65.
- Njwe, R.M., 1992. Protein requirements of Cameroonian Dwarf goats. World Rev. Anim. Prod. 27, 23–29.
- Osafo, E.L.K., Okai, D.B., Buadu, M.K., Hammond, F., 1996. The effect of gliricidia supplementation on feed intake and liveweight change of West African dwarf goats fed coffee-pulp containing diet. In: Holst, P.J. (Ed.), Proceedings of the VI International Conference on Goats. International Academic Publishers, Beijing, China, pp. 630–632.
- Osuji, P.O., 1987. Intensive feeding systems for goats in Latin America and the Carribean. In: Santana, O.P., da Silva, A.G., Foote, W.C. (Eds.), Proceedings of the IV International Conference on Goats. Departamento de Difusao de Tecnologia, Brasilia, Brazil, pp. 1077–1108.
- Pfeffer, E., Rodehutscord, M., 1998. Body chemical composition and utilization of dietary energy by male Saanen kids fed either milk to satiation or

solid complete feeds with two proportions of straw. J. Agric. Sci. (Camb.) 131, 487–495.

- Phimphachanhvongsod, V., Ledin, I., 2002. Performance of growing goats fed *Panicum maximum* and leaves of *Gliricidia sepium*. Asian-Aust. J. Anim. Sci. 15, 1585–1590.
- Prieto, I., Goetsch, A.L., Banskalieva, V., Cameron, M., Puchala, R., Sahlu, T., Dawson, L.J., Coleman, S.W., 2000. Effects of dietary protein concentration on postweaning growth of Boer crossbred and Spanish goat wethers. J. Anim. Sci. 78, 2275–2281.
- Qi, K., Lu, C.D., Owens, F.N., 1993. Sulfate supplementation of growing goats: effects on performance, acid-base balance, and nutrient digestibilities. J. Anim. Sci. 70, 1579–1587.
- Rengsirikul, B., Sae Nai, P., 1991. The use of palm oil meal with urea-treated rice straw in postweaning goat rations. In: Saithanoo, S., Norton, B.W. (Eds.), Goat Production in the Asian Humid Tropics. The Australian International Development Assistance Bureau, Hat Yai, Thailand, pp. 154– 163.
- Rodriguez, W.E., Murillo, B., Velez, M., 1992a. *Gliricidia sepium* leaves as a forage. I. Intake and digestibility. In: Lokeshwar, R.R. (Ed.), Recent Advances in Goat Production. Proceedings of the V International Conference on Goats. Nutan Printers, New Delhi, India, pp. 761–764.
- Rodriguez, W.E., Velez, M., Esnaola, M.A., 1992b. *Gliricidia sepium* leaves as a forage. 2. Kids and lamb fattening. In: Lokeshwar, R.R. (Ed.), Recent Advances in Goat Production. Proceedings of the V International Conference on Goats. Nutan Printers, New Delhi, India, pp. 765–772.
- Saikia, G., Baruah, K.K., Buragohain, S.C., Saikia, B.N., Pathak, N.N., 1995. Feed intake, utilization of nutrients and growth of Assamese × Beetal goats fed three levels of energy. Small Rumin. Res. 15, 279–282.
- Samanta, A.K., Singh, K.K., Das, M.M., Maity, S.B., Kundu, S.S., 2003. Effect of complete feed block on nutrient utilisation and rumen fermentation in Barbari goats. Small Rumin. Res. 48, 95–102.
- Shelton, M., Thompson, P.V., 1976. Influence of protein level and monensin on performance of male kid goats fed in drylot. In: Sheep Goat Wool Mohair. Texas Agricultural Experimental Station, San Angelo, TX, PR-3395, pp. 26–27.

- Sooden-Karamath, S., Youssef, F.G., 1999. Effect of monensin, avoparcin and grass supplementation on utilization of urea-treated rice straw by sheep and goats. Small Rumin. Res. 33, 201–211.
- Soto-Navarro, S.A., Goetsch, A.L., Sahlu, T., Puchala, R., 2004. Effect of level and source of supplemental protein in a concentrate-based diet on growth performance of Boer × Spanish wether goats. Small Rumin. Res. 51, 101–106.
- Souri, M., Galbraith, H., Scaife, J.R., 1998. Comparison of the effect of genotype and protected methionine supplementation on growth, digestive characteristics and fibre yield in cashmere-yielding and Angora goats. Anim. Sci. 66, 217– 223.
- Titi, H.H., 2003. Replacing soybean meal with sunflower meal with or without fibrolytic enzymes in fattening diets of goat kids. Small Rumin. Res. 48, 45–50.
- Vadiveloo, J., 1988. Performance of young indigenous and crossbred goats fed forages supplemented with palm oil mill effluent. Small Rumin. Res. 1, 369–379.
- Van Eys, J.E., Mathius, I.W., Pongsapan, P., Johnson, W.L., 1986. Foliage of the tree legumes gliricidia, leucaena, and sesbania as supplement to napier grass diets for growing goats. J. Agric. Sci. (Camb.) 107, 227–233.
- van Hao, N., Ledin, I., 2001. Performance of growing goats fed *Gliricidia maculata*. Small Rumin. Res. 39, 113–119.
- Verma, A.K., Sastry, V.R.B., Agrawal, D.K., 1995. Feeding of water washed neem (*Azadirachta indica*) seed kernel cake to growing goats. Small Rumin. Res. 15, 105–111.
- Virk, A.S., Khatta, V.K., Tewatia, B.S., Gupta, P.C., 1994. Effect of formaldehyde-treated faba beans (*Vicia faba* L.) on nutrient utilization and growth performance of goat kids. Small Rumin. Res. 14, 19–23.
- Yami, A., Litherland, A.J., Davis, J.J., Sahlu, T., Puchala, R., Goetsch, A.L., 2000. Effects of dietary level of *Leucaena leucocephala* on performance of Angora and Spanish doelings. Small Rumin. Res. 38, 17–27.
- Yan, T., Cook, J.E., Gibb, M.J., Ivings, W.E., Treacher, T.T., 1993. The effects of quantity and duration of milk feeding on the intake of concentrates and

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growth of castrated male Saanen kids to slaughter. Anim. Prod. 56, 327–332.

Zemmelink, G., Tolkamp, B.J., Ogink, N.M.W., 1991. Energy requirements for maintenance and gain of West African Dwarf goats. Small Rumin. Res. 5, 205–215.

#### References

- AFRC, 1992. Nutritive requirements of ruminant animals: protein. Nutr. Abstr. Rev. 62, 787–835.
- AFRC, 1993. Energy and Protein Requirements of Ruminants. CAB International, Wallingford, UK, pp. 1–122.
- AFRC, 1998. The Nutrition of Goats. CAB International, New York, NY, pp. 7–64.
- Ahn, J.H., Jo, I.H., Lee, J.S., 2002. The use of apple pomace in rice straw based diets of Korean native goats (*Capra hircus*). Asian-Aust. J. Anim. Sci. 15, 1599–1605.
- Ainslie, S.J., Fox, D.G., Perry, T.C., Ketchen, D.J., Barry, M.C., 1993. Predicting amino acid adequacy of diets fed to Holstein steers. J. Anim. Sci. 71, 1312–1319.
- ARC, 1980. The Nutrient Requirements of Ruminant Livestock. Commonwealth Agricultural Bureaux, Slough, UK, p. 17.
- Ash, A.J., 1990. The effect of supplementation with leaves from the leguminous trees *Sesbania grandiflora*, *Albizia chinensis* and *Gliricidia sepium* on the intake and digestibility of guinea grass hay by goats. Anim. Feed Sci. Technol. 28, 225–232.
- Baah, J., Tait, R.M., Tuah, A.K., McAllister, T.A., Bae, H.D., Cheng, K.J., 1999. Examination of microbial degradation of *Ficus exasperata* leaves and cassava peels by in situ incubation and scanning electron microscopy. Anim. Feed Sci. Technol. 77, 213–228.
- Brown, M.S., Galyean, M.L., Duff, G.C., Hallford, D.M., Soto-Navarro, S.A., 1998. Effects of degree of processing and nitrogen source and level on starch availability and in vitro fermentation of corn and sorghum grain. Prof. Anim. Sci. 14, 83–94.
- Byers, F.M., 1982. Protein growth and turnover in cattle: systems for measurement and biological limits. In: Owens, F.N. (Ed.), Protein Requirements for Cattle. Oklahoma State University, MP-109, pp. 141–165.
- Chatterjee, S., Hadi, A.S., Price, B., 2000. Regression Analysis by Example, 3rd ed. Wiley, New York, NY, pp. 285–312.
- Fleischer, J.E., Sottie, E.T., Amaning-Kwarteng, K., 1998. Chemical composition and rumen degradability of protein of browse and shrubs fed to sheep in Ghana. Ghana J. Agric. Sci. 38, 93–98.
- Hadjipanayiotou, M., 2002. Replacement of soybean meal and barley grain by chickpeas in lamb and kid fattening diets. Anim. Feed Sci. Technol. 96, 103–109.
- Ibrahim, M.N.M., Tamminga, S., Zemmelink, G., 1989. Effect of urea treatment on rumen degradation characteristics of rice straws. Anim. Feed Sci. Technol. 24, 83–95.

- INRA, 1989. In: Jarrige, R. (Ed.), Ruminant Nutrition: Recommended Allowances and Feed Tables. INRA, Paris, France, pp. 175–176.
- Kaitho, R.J., Umunna, N.N., Nsahlai, I.V., Tamminga, S., van Bruchem, J., 1998. Nitrogen in browse species: ruminal degradability and post-ruminal digestibility measured by mobile nylon bag and in vitro techniques. J. Sci. Food Agric. 76, 488– 498.
- Kalbande, V.H., Thomas, C.T., 1997. Ruminal degradability evaluations of green forages by nylon-bag technique. Indian J. Anim. Sci. 67, 231–233.
- Liu, J.X., Yao, J., Yan, B., Yu, J.Q., Shi, Z.Q., Wang, X.Q., 2000. The nutritional value of mulberry leaves and their use as supplement to growing sheep fed ammoniated rice straw. In: Sanchez, M., Speedy, A. (Eds.), Proceedings of the FAO Electronic Conference on "Mulberry for Animal Production". The Feed Resources Group of FAO, Website, pp. 1–13.
- Luo, J., Goetsch, A.L., Moore, J.E., Johnson, Z.B., Sahlu, T., Ferrell, C.L., Galyean, M.L., Owens, F.N., 2004. Prediction of endogenous urinary nitrogen of goats. Small Rumin. Res. 53, 293–308.
- Mgheni, D.M., Hvelplund, T., Weisbjerg, M.R., 1996. Rumen degradability of dry matter and protein in tropical grass and legume forages and their protein values expressed in the AAT-PBV protein evaluation system. In: Ndikumana, J., de Leeuw, P. (Eds.), Sustainable Feed Production and Utilization of Smallholder Livestock Enterprises in Sub-Saharan Africa. Proceedings of the Second African Feed Resources Network (AFRNET), African Feed Resources Network, Nairobi, Kenya, pp. 95–100.
- Michalet-Doreau, B., Nozière, P., 1998. Validation of in situ nitrogen degradation measurements: comparative proteolytic activity of solid-adherent microorganisms isolated from rumen content and nylon bags containing various feeds. Anim. Feed Sci. Technol. 70, 41–47.
- Mmbengwa, V.M., Schwalbach, L.M., Greyling, J.P.C., Fair, M.D., 2000. Milk production potential of South African Boer and Nguni goats. S. Afr. J. Anim. Sci. 30 (Suppl. 1), 76–77.
- Moore, J.E., Goetsch, A.L., Luo, J., Owens, F.N., Galyean, M.L., Johnson, Z.B., Sahlu, T., Ferrell, C.L., 2004. Prediction of fecal crude protein excretion of goats. Small Rumin. Res. 53, 275– 292.
- Negi, S.S., Singh, B., Makkar, H.P.S., 1988. An approach to the determination of rumen degradability of nitrogen in low-grade roughages and partition of nitrogen therein. J. Agric. Sci. (Camb.) 111, 487–494.
- Ngwa, A.T., Nsahlai, I.V., Bonsi, M.L.K., 2001. The effect of feeding pods of multipurpose trees (MPTs) on the degradability of dry matter and cell wall constituents of maize stover and alfalfa incubated in the rumen of sheep. J. Sci. Food Agric. 81, 1235–1243.
- NRC, 1981. Nutrient Requirements of Goats: Angora, Dairy, and Meat Goats in Temperate and Tropical Countries. National Academy Press, Washington, DC, pp. 26–48.
- NRC, 1984. Nutrient Requirements of Beef Cattle, 6th ed. National Academy Press, Washington, DC, pp. 47–61.
- NRC, 1985. Ruminant Nitrogen Usage. National Academy Press, Washington, DC, pp. 61–63.

- NRC, 2000. Nutrient Requirements of Beef Cattle, 7th ed. (update). National Academy Press, Washington, DC, pp. 16–21, 191–228.
- NRC, 2001. Nutrient Requirements of Dairy Cattle, 7th ed. National Academy Press, Washington, DC, pp. 67–69.
- Nsahlai, I.V., Bryant, M.J., Umunna, N.N., 1999. The utilization of barley straw by steers: effects of replacing urea with protein, source of protein and quantity of rumen degradable nitrogen on straw degradation, liquid and particle passage rates and intake. J. Appl. Anim. Res. 16, 129–146.
- Nsahlai, I.V., Goetsch, A.L., Luo, J., Johnson, Z.B., Moore, J.E., Sahlu, T., Ferrell, C.L., Galyean, M.L., Owens, F.N., 2004. Metabolizable protein requirements of lactating does. Small Rumin. Res. 53, 327–337.
- Oldham, J.D., 1987. Efficiencies of amino acid utilization. In: Jarrige, R., Alderman, G. (Eds.), Feed Evaluation and Protein Requirement Systems for Ruminants. Commission of the European Communities, Luxembourg, Belgium, pp. 171–186.
- Ørskov, E.R., 1980. Degradability of protein supplements and utilization of undegraded protein by high-producing dairy cows. In: Haresign, W. (Ed.), Recent Advances in Animal Nutrition. Butterworths, London, UK, pp. 85–98.
- Ørskov, E.R., DeB Hovell, F.D., Mould, F., 1980. The use of the nylon bag technique for the evaluation of feedstuffs. Trop. Anim. Prod. 5, 195–213.
- Ørskov, E.R., MacLeod, N.A., 1982. Validation and application of new principles of protein evaluation for ruminants. In:

Miller, E.L., Pike, I.H., Van Es, A.J.H. (Eds.), Protein Contribution of Feedstuffs for Ruminants: Application to Feed Formulation. Butterworth Scientific, London, UK, pp. 76–85.

- Ørskov, E.R., Shand, W.J., 1997. Use of the nylon bag technique for protein and energy evaluation and for rumen environment studies in ruminants. Livest. Res. Rural Dev. 9, 9–12.
- Sampath, K.T., Rao, A.S., Sampath, S.R., 1989. Ruminal protein degradability of certain feedstuffs determined by nylon-bag technique. Indian J. Anim. Sci. 59, 1304–1307.
- SAS, 1990. SAS User's Guide: Statistics, 6th ed. SAS Institute Inc., Cary, NC.
- SCARM, 1994. Feeding Standards for Australian Livestock: Ruminants. CSIRO, Victoria, Australia, pp. 18–21, 51–54, and 107–109.
- Snedecor, G.W., Cochran, W.G., 1978. Statistical Methods, 6th ed. The Iowa State University Press, Ames, IA, pp. 419–446.
- Soto-Navarro, S.A., Goetsch, A.L., Sahlu, T., Puchala, R., 2004. Effect of level and source of supplemental protein in a concentrate-based diet on growth performance of Boer×Spanish wether goats. Small Rumin. Res. 51, 101–106.
- Vadiveloo, J., 2000. Nutritional properties of the leaf and stem of rice straw. Anim. Feed Sci. Technol. 83, 57–65.
- Wilkerson, V.A., Klopfenstein, T.J., Britton, R.A., Stock, R.A., Miller, P.S., 1993. Metabolizable protein and amino acid requirements of growing cattle. J. Anim. Sci. 71, 2777– 2784.